

Term Contract for Provision of Sampling and Analyzing of Wastewater and Sludge Samples for Various Sewage Treatment Facilities and Marine Water Samples in Urban Area, Lantau and Outlying Islands to the Drainage Services Department

Whole Effluent Toxicity Test (WETT) at SCISTW

Report for the Month of January 2023

Contract No. : DE/2020/02

Applicant : DRAINAGE SERVICES DEPT. - DIVISION 2

Address : STONECUTTERS ISLAND SEWAGE

TREATMENT WORKS, NGONG SHUEN CHAU,

KOWLOON, HONG KONG

Application No. : L0000379(8)

Report No. : A00000733(2)

Report Issued Date : 03 Feb 2023

For and on behalf of

CMA Industrial Development Foundation Limited

Authorized Signature:

Lau Yan Kin Senior Manager



Co	onte	nts	2
1	Intr	roduction	4
	1.1	Background	4
	1.2	Testing laboratory and investigator	4
	1.3	Sample	4
	1.4	Test species	4
	1.5	Test protocols	4
2	Rep	oort on Amphipod Acute Toxicity Test	5
	2.1	Samples storage and pretreatment	6
	2.2	Test organism	6
	2.3	Summary of test conditions	6
	2.4	Test results	7
	2.5	Summary of water quality parameters monitoring during test	8
	2.6	LC ₅₀ for the amphipod <i>Melita longidactyla</i> and test acceptability	8
3	Rep	oort on Fish Acute Toxicity Test	9
	3.1	Samples storage and pretreatment	10
	3.2	Test organism	10
	3.3	Summary of test conditions	10
	3.4	Test results	11
	3.5	Summary of water quality parameters monitoring during test	12
	3.6	LC ₅₀ for the fish Lutjanus malabaricus and test acceptability	12
4	Rep	ort on Barnacle Larvae Acute Toxicity Test	13
	4.1	Samples storage and pretreatment	14
	4.2	Test organism	14
	4.3	Summary of test conditions	14
	4.4	Test results	15
	4.5	Summary of water quality parameters monitoring during test	16
	4.6	LC ₅₀ for the barnacle larvae <i>Balanus amphitrite</i> and test acceptability	16
5	Rep	oort on Diatom Growth Inhibition Test (Chronic toxicity test)	17
	5.1	Samples storage and pretreatment	18
	5.2	Test organism	18
	5.3	Summary of test conditions	18
	5.4	Test results	19
	5.5	Summary of water quality parameters monitoring during test	20
	5.6	IC ₅₀ for the diatom <i>Skeletonema costatum</i> and test acceptability	20
6	Rep	ort on Shrimp Acute Toxicity Test	21
	6.1	Samples storage and pretreatment	22
	6.2	Test organism	22
	6.3	Summary of test conditions	22
	6.4	Test results	2 3
	6.5	Summary of water quality parameters monitoring during test	24



6.6 LC ₅₀ for the shrimp <i>Metapenaeus ensis</i> and test acceptability	24
7 Conclusion	25-26
Appendix A Monitoring Data for Amphipod Acute Toxicity Test	27-29
Appendix B Monitoring Data for Fish Acute Toxicity Test	30-32
Appendix C Monitoring Data for Barnacle Larvae Acute Toxicity Test	33-35
Appendix D Monitoring Data for Diatom Growth Inhibition Test	36-38
Appendix E Monitoring Data for Shrimp Acute Toxicity Test	39-41



1. Introduction

1.1. Background

The whole effluent toxicity tests (WETT) were carried out under the requirements of Drainage Service Department (DSD).

1.2. Testing laboratory and investigator

Name of testing laboratory: Research Centre for the Oceans and Human Health Principle investigator: Prof. Wen-Xiong WANG

1.3. Sample

A 24-hour flow-weighted composite effluent sample was collected from Stonecutters Island Sewage Treatment Works (SCISTW) on January 14, 2023. Effluent sample was shipped immediately to the testing laboratory on the same day of collection. Toxicity testings started immediately after the arrival of samples.

1.4. <u>Test species</u>

The following test species were included in the WETT:

- Amphipod (Melita longidactyla)
- Fish (*Lutjanus malabaricus*)
- Barnacle larvae (Balanus amphitrite)
- Diatom (Skeletonema costatum)
- Shrimp (*Metapenaeus ensis*)

1.5. <u>Test protocols</u>

The WETT testing methods and procedures follow those documented in "Consultancy Study on Fisheries and Marine Ecological Criteria for Impact Assessment-Final Report" commissioned by Agriculture, Fisheries and Conservation Department (AFCD), as indicated in tender addendum No. 1 by Drainage Services Department (DSD).



2. Report on Amphipod Acute Toxicity Test



Test report

2.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

2.2. Test organism

Species Amphipod (Melita longidactyla).

Source: Collected from local coastal waters from Sai Kung

Size/age: 0.5-0.7 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature:

22±1°C, salinity: 30‰) at least 48 hours in the laboratory

prior to test. Fed with green algae *Ulva lactuca*.

2.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 14/1/2023-16/1/2023

Dilution seawater source: Seawater collected from a pristine site in Clear Water

Bay, Sai Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

Testing temperature: 22±1 °C Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 150 mL glass flask

Feeding: None

Number of organisms per

10

replicate:

Replicate number: 4

Volume of test medium: 100 mL

Aeration: Moderate, around 100 bubbles/min

Reference toxicant: CdCl₂

Positive control: 48-h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



2.4. <u>Test results</u>

Table 1. Survival of amphipods after 48 hours.

Treatment	Effluent	Number of living amphipods after 48 hour (individuals)					
	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	10	10	8	8	9.00	1.15
Salinity control	0	10	9	9	8	9.00	0.82
Concentration 1	6.5	7	9	8	8	8.00	0.82
Concentration 2	12.5	6	8	7	7	7.00	0.82
Concentration 3	25	5	6	4	5	5.00	0.82
Concentration 4	50	0	2	0	3	1.25	1.50
Concentration 5	100	0	0	0	0	0.00	0.00

Table 2. Survival percentage of amphipods after 48 hours.

	Effluent		Percentage	of living amr	hipods after 4	8 hour (%)	
Treatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	\ /	100	100	80	80	90.00	11.55
Salinity control		100	90	90	80	90.00	8.17
Concentration 1	6.5	70	90	80	80	80.00	8.17
Concentration 2	***	60	80	70	70	70.00	8.17
Concentration 3	25	50	60	40	50	50.00	8.17
Concentration 4	50	0	20	0	30	12.50	15.00
Concentration 5	100	0	0	0	0	0.00	0.00



2.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during amphipod acute toxicity test.

		Ef	fluent concer	ntration (%)			
Water quality parameters	Negative	Salinity	6.5	12.5	25	50	100
	control	control	0.5	12.5	23	30	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L-1)	6.8-7.2	6.8-7.1	6.9-7.1	6.9-7.1	6.8-7.1	6.9-7.0	6.8-7.0
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
рН	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.0	7.8-8.1	7.8-7.9
Ammonia-N (start/end, mg L-1)	0.87/0.07	0.95/<0.01	3.18/1.86	6.34/3.10	8.42/5.77	13.9/11.4	26.4/23.8
Total Sulphide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	0.13/<0.1
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Total suspended solids (start/end, mg L-1)	50/<2	60/<2	70/5	77/6	80/11	89/26	110/41

2.6. LC₅₀ for the amphipod *Melita longidactyla* and test acceptability

Table 4. LC_{50} for the amphipods and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	28.3%	NA
Negative control survival	90%	≥90%
Reference toxicant 48-h acute test	0.96 mg L ⁻¹	1.25±0.15 mg L ⁻¹
95% of confidence range of reference toxicant test	0.7-1.33 mg L ⁻¹	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: ±1 °C
Dissolved oxygen concentration	>6.8 mg L ⁻¹	>4 mg L-1

NA: Not applicable



3. Report on Fish Acute Toxicity Test



Test report

3.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

3.2. <u>Test organism</u>

Species Fish (Lutjanus malabaricus)

Source: Purchased from contracted fish farm

Size/age: 2-3 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature:

22±1°C, salinity: 30‰) at least 48 hours in laboratory prior to test. Fed with fresh shrimp purchased from local

market.

3.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 14/1/2023-16/1/2023

Dilution seawater source: Seawater collected from a pristine site in Clear Water

Bay, Sai Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 5 µm filtration bag

Testing temperature: 22±1 °C Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 20 L tank

Feeding: None

Number of organisms per replicate: 20

Replicate number: 4
Volume of test medium: 20 L

Aeration: Moderate, with air stone

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



3.4. <u>Test results</u>

Table 1. Survival of fish after 48 hours.

Treatment	Effluent		Number of	living fish af	ter 48 hour (in	dividuals)	
Treatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	20	20	20	20	20.00	0.00
Salinity control	0	20	20	20	20	20.00	0.00
Concentration 1	6.5	20	20	20	20	20.00	0.00
Concentration 2	12.5	20	19	20	18	19.25	0.96
Concentration 3	25	20	20	20	20	20.00	0.00
Concentration 4	50	19	19	18	17	18.25	0.96
Concentration 5	100	18	18	17	16	17.25	0.96

Table 2. Survival percentage of fish after 48 hours.

Treatment	Effluent	Percentage of living fish after 48 hour (%)					
reatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative contro	1 0	100	100	100	100	100.00	0.00
Salinity control	0	100	100	100	100	100.00	0.00
Concentration 1	6.5	100	100	100	100	100.00	0.00
Concentration 2	12.5	100	95	100	90	96.25	4.79
Concentration 3	25	100	100	100	100	100.00	0.00
Concentration 4	50	95	95	90	85	91.25	4.79
Concentration 5	100	90	90	85	80	86.25	4.79



3.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during fish acute toxicity test.

Matan quality narramators		Eff	luent concer	ntration (%)			
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L-1)	6.8-7.0	6.9-7.1	6.9-7.1	6.8-7.1	6.8-7.0	6.8-7.1	6.7-7.0
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.9-8.0	7.8-8.0	7.9-8.0	7.7-8.1	7.8-8.1	7.8-8.0	7.9-8.0
Ammonia-N (start/end, mg L-1)	< 0.01	< 0.01	1.95/1.45	3.09/3.06	6.07/5.42	12.9/10.3	23.6/23.0
Total Sulphide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	0.19/< 0.1
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Total suspended solids (mg L-1)	< 2	< 2	3/2	6/<2	10/3	16/6	41/8

3.6. <u>LC₅₀ for the fish Lutjanus malabaricus</u> and test acceptability

Table 4. LC₅₀ for the fish and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	Cannot be calculated ^a	NA
Negative survival	100%	>90%
Reference toxicant 48-h acute test	14.1 mg L ⁻¹	14.6±1.78 mg L ⁻¹
95% of confidence range of reference toxicant	10.7-18.2 mg L ⁻¹	NI A
test	_	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: ± 1 °C
Dissolved oxygen concentration	>6.6 mg L ⁻¹	>4 mg L ⁻¹

a: The mortalities in all concentration groups were less than 50% and thus LC₅₀ cannot be calculated.

NA: Not applicable



4. Report on Barnacle Larvae Acute Toxicity Test



Test report

4.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 μ m membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

4.2. Test organism

Species Barnacle larvae (Balanus amphitrite).

Source: Introduced from adult barnacles collected from Sai Kung

Size/age: Stage II

Acclimatization: Acclimatized in fully aerated seawater held in 500 mL

glass beaker (temperature: 22±1°C, salinity: 30%) for at least 24 hours in laboratory prior to test. Fed with

diatom Chaetoceros gracilis.

4.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 14/1/2023-16/1/2023

Dilution seawater source: Seawater collected from a pristine site in Clear Water

Bay, Sai Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 µm membrane

Testing temperature: 22±1 °C
Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 50 mL glass beaker

Feeding: None

Number of organisms per

replicate:

20

Replicate number: 4
Volume of test medium: 20 mL

Aeration: Moderate, around 100 bubbles/min

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



4.4. <u>Test results</u>

Table 1. Survival of barnacle larvae after 48 hours.

Treatment	Effluent	Number of living barnacle larvae after 48 hour (individuals)					
	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	20	20	20	20	20.00	0.00
Salinity control	0	20	20	20	20	20.00	0.00
Concentration 1	6.5	20	20	20	20	20.00	0.00
Concentration 2	12.5	15	13	13	14	13.75	0.96
Concentration 3	25	10	14	12	12	12.00	1.63
Concentration 4	50	12	10	10	12	11.00	1.15
Concentration 5	100	0	0	0	0	0.00	0.00

Table 2. Survival percentage of barnacle larvae after 48 hours.

	0						
Treatment	Effluent		Percentage of	f living barna	cle larvae after	48 hour (%)	
Heatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	100	100	100	100	100.00	0.00
Salinity control	0	100	100	100	100	100.00	0.00
Concentration 1	6.5	100	100	100	100	100.00	0.00
Concentration 2	12.5	75	65	65	70	68.75	4.79
Concentration 3	25	50	70	60	60	60.00	8.16
Concentration 4	50	60	50	50	60	55.00	5.77
Concentration 5	100	0	0	0	0	0.00	0.00



4.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during barnacle larvae acute toxicity test

	8	Effl	uent concer	ntration (%)			
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L-1)	6.8-6.9	6.8-7.0	6.8-7.0	6.8-6.9	6.8-7.0	6.8-7.1	6.8-6.9
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0
рН	7.8-8.0	7.8-8.0	7.9-8.0	7.9-8.0	7.8-7.9	7.7-7.9	7.7-7.8
Ammonia-N (start/end, mg L-1)	0.01/<0.01	< 0.01	1.23/0.97	2.53/2.00	5.83/4.44	10.5/8.76	22.9/17.4
Total Sulphide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	0.15/<0.1
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Total suspended solids (mg L-1)	< 2	< 2	3/<2	4/<2	11/<2	17/< 2	41/3

4.6. <u>LC₅₀ for the barnacle larvae Balanus amphitrite and test acceptability</u>

Table 4. LC₅₀ for the barnacle larvae and test acceptability

Parameter	Value	Control limit
Calculated LC ₅₀	31.8%	NA
Negative survival	100%	>90%
Reference toxicant 48-h acute test	1.12 mg L ⁻¹	1.04±0.11 mg L ⁻¹
95% of confidence range of reference toxicant	0.89-1.34 mg L ⁻¹	NA
test:		1 1/2 1
Daily temperature variation	<0.5 °C	Average daily temperature variation: ± 1 °C
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L ⁻¹

NA: Not applicable



5. Report on Diatom Growth Inhibition Test (Chronic toxicity test)



Test report

5.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 μ m membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

5.2. Test organism

Species Diatom (Skeletonema costatum)

Grown from laboratory culture obtained from Coastal

Source: Marine Lab, Hong Kong University of Science and

Technology

Size/age: Log growth phase

Grown in 250 mL glass flask (temperature: 22±1°C,

Acclimatization: salinity: 30%, 3000 lux) for at least two weeks prior to

test.

5.3. Summary of test conditions

Type of test: Static

Duration: 7 days, 14/1/2023-21/1/2023

Dilution seawater source: Seawater collected from a pristine site in Clear Water

Bay, Sai Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

Testing temperature: 22±1 °C

Lighting: 12 h light/12 h dark cycle, 3000±500 lux

Salinity: 30%

Testing chamber: Pre-cleaned 100 mL glass beaker

Initial cell density: $(5.0\pm0.4)\times10^4$ cell mL⁻¹

Replicate number: 4

Volume of test medium: 25 mL Aeration: None Reference toxicant: CdCl₂

Positive control: 7-day IC₅₀ toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



5.4. Test results

Table 1. Cell density of diatom *Skeletonema costatum* at the beginning and end of growth inhibition test. Initial cell density: (5.0±0.4)×10⁴ cell mL⁻¹.

Treatment	Effluent		Cell densi	ty after 7-day	growth (×106	cell mL-1)	
	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	1.78	1.88	1.69	1.73	1.77	0.08
Salinity control	0	1.85	1.90	1.72	1.80	1.82	0.08
Concentration 1	2.5	2.52	2.60	2.43	2.33	2.47	0.12
Concentration 2	5	3.35	3.58	3.47	3.65	3.51	0.13
Concentration 3	10	2.25	2.28	2.50	2.43	2.37	0.12
Concentration 4	25	2.10	2.07	2.14	2.00	2.08	0.06
Concentration 5	50	0.13	0.15	0.30	0.08	0.17	0.09
Concentration 6	100	0.05	0.03	0.00	0.17	0.06	0.07

Table 2. Growth rate of *Skeletonema costatum* within 7 days.

Treatment	Effluent	7-day average growth rate (d-1)						
reatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	0.51	0.52	0.50	0.51	0.51	0.01	
Salinity control	0	0.52	0.52	0.51	0.51	0.51	0.01	
Concentration 1	2.5	0.56	0.56	0.55	0.55	0.56	0.01	
Concentration 2	5	0.60	0.61	0.61	0.61	0.61	0.01	
Concentration 3	10	0.54	0.55	0.56	0.55	0.55	0.01	
Concentration 4	25	0.53	0.53	0.54	0.53	0.53	0.00	
Concentration 5	50	0.14	0.16	0.26	0.07	0.15	0.08	
Concentration 6	100	0.00	0.00	0.00	0.17	0.04	0.09	



5.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during diatom growth inhibition test

Maton quality namatons			Efflue	ent concentra	tion (%)			
Water quality parameters	Negative control	Salinity control	2.5	5.0	10	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L-1)	6.8-8.2	6.8-8.2	6.8-8.2	6.8-8.2	6.8-8.3	6.8-8.2	6.8-7.9	6.8-7.3
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pH	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.0
Ammonia-N (start/end, mg L ⁻¹)	0.01/<0.01	< 0.01	2.49/<0.01	2.33/<0.01	5.40/<0.01	7.72/<0.01	11.2/5.09	22.4/19.5
Total Sulphide (start/end, mg L-1)	< 0.1	< 0.1	0.12/< 0.1	0.16/< 0.1	0.18/< 0.1	0.22/< 0.1	0.23/< 0.1	0.26/0.24
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Total suspended solids (mg L-1)	< 2	< 2	3/<2	6/4	9/5	15/10	19/14	47/30

5.6. IC₅₀ for the diatom *Skeletonema costatum* and test acceptability

Table 4. IC₅₀, none observed effect concentration (NOEC) for the diatom and test acceptability

Parameter	Value	Control limit
Calculated IC ₅₀	36.5%	NA
None observed effect concentration (NOEC)	25%	-
Reference toxicant 7-day test:	0.17 mg L ⁻¹	$0.13\pm0.02~{ m mg}~{ m L}^{ ext{-}1}$
95% of confidence range of reference toxicant test	0.11-0.26 mg L ⁻¹	NA
Temperature variation	<0.5 °C	Average daily temperature variation: ± 1 °C

NA: Not applicable



6. Report on Shrimp Acute Toxicity Test



Test report

6.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to 25‰ and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

6.2. Test organism

Species Shrimp (Metapenaeus ensis)

Source: Purchased from contracted fish dealer

Size/age: 5-7 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature:

22±1°C, salinity: 25‰) at least 48 hours in the laboratory

prior to test. Fed with commercial shrimp feeds.

6.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 14/1/2023-16/1/2023

Dilution seawater source: Seawater collected from a pristine site in Clear Water

Bay, Sai Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

10

Testing temperature: 22±1 °C Lighting: Continuous

Salinity: 25‰

Testing chamber: Pre-cleaned 20 L tank

Feeding: None

Number of organisms per

replicate:

Replicate number: 4

Replicate number: 4

Volume of test medium: 10 L Aeration: Moderate, with air stone

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 25‰



6.4. Test results

Table 1. Survival of shrimps after 48 hours.

Treatment	Effluent	Number of living shrimps after 48 hour (individuals)						
Heatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	10	9	9	8	9.00	0.82	
Salinity control	0	9	8	9	9	8.75	0.50	
Concentration 1	6.5	8	6	8	8	7.50	1.00	
Concentration 2	12.5	7	8	8	6	7.25	0.96	
Concentration 3	25	9	6	6	8	7.25	1.50	
Concentration 4	50	6	7	7	6	6.50	0.58	
Concentration 5	100	6	5	5	6	5.50	0.58	

Table 2. Survival percentage of shrimps after 48 hours.

Tuontun on t	Effluent	Percentage of living shrimps after 48 hour (%)						
Treatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	100	90	90	80	90.00	8.16	
Salinity control	0	90	80	90	90	87.50	5.00	
Concentration 1	6.5	80	60	80	80	75.00	10.00	
Concentration 2	12.5	70	80	80	60	72.50	9.57	
Concentration 3	25	90	60	60	80	72.50	15.00	
Concentration 4	50	60	70	70	60	65.00	5.77	
Concentration 5	100	60	50	50	60	55.00	5. <i>77</i>	



6.5. Summary of water quality parameters monitoring during test.

Table 3. Summary of water quality parameters during shrimp acute toxicity test.

Maken quality nanamakana	Effluent concentration (%)								
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100		
Salinity (‰)	25.0	25.0	25.0	25.0	25.0	25.0	25.0		
Dissolved oxygen (mg L-1)	6.8-6.9	6.9-7.0	6.9-7.0	6.9-7.0	6.9-7.1	6.8-7.0	6.8-7.0		
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0		
pH	7.9-8.0	7.9-8.0	7.9-8.0	7.9-7.9	7.9-8.0	7.7-7.9	7.7-8.0		
Ammonia-N (start/end, mg L-1)	1.67/<0.01	1.80/<0.01	2.83/2.37	5.04/4.48	4.15/4.07	14.5/14.0	24.5/23.9		
Total Sulphide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	0.24/< 0.1		
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02		
Total suspended solids (start/end, mg L-1)	< 2	2/< 2	6/<2	6/4	7/2	27/6	37/9		

6.6. LC₅₀ for the shrimp Metapenaeus ensis and test acceptability

Table 4. LC₅₀ for the *Metapenaeus ensis* and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	Cannot be calculated ^a	NA
Negative control survival	90%	>90%
Reference toxicant 48-h acute test	6.15 mg L ⁻¹	$5.19\pm0.51~{ m mg}~{ m L}^{-1}$
95% of confidence range of reference toxicant test	5.04-7.41 mg L ⁻¹	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: ±1 °C
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L-1
	1 -00/ 11 -0	

a: The mortalities in all concentration groups were less than 50% and thus LC₅₀ cannot be calculated.

NA: Not applicable



7. Conclusion



Table 1. Comparison of measured toxicity values with the target toxicity levels.

Test species	Measured LC ₅₀ /IC ₅₀ /NOEC	Target toxicity level
Amphipod Melita longidactyla	28.3%	≥7.1%
Fish Lutjanus malabaricus	Cannot be calculated because animals displayed very high survival rate (>50%) at the highest tested effluent concentration (100%)	≥7.1%
Barnacle larvae Balanus amphitrite	31.8%	≥7.1%
Diatom Skeletonema costatum	36.5%	-
Diatom Skeletonema costatum	25% (NOEC)	≥0.51%
Shrimp Metapenaeus ensis	Cannot be calculated because animals displayed very high survival rate (>50%) at the highest tested effluent concentration (100%)	≥7.1%

Conclusion: all the measured values met the target toxicity levels as indicated in the EM&A Manual.

Cannot be calculated: The mortalities in all concentration groups (even at the highest effluent concentration) were less than 50% and thus LC_{50} cannot be calculated.



Appendix A

Monitoring Data for Amphipod Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in amphipod acute toxicity test.

Concentration treatment (%)		Dissolve	d oxygen	(mg L-1)		рН				
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48h
Negative control	6.9	7.2	7.2	6.9	6.8	7.9	7.9	8.1	8.1	8.0
Salinity control	6.8	6.9	7.1	7.1	7.0	7.9	8.0	8.1	8.1	8.1
6.5	6.9	7.0	7.0	7.0	7.1	8.0	7.9	8.1	8.1	8.1
12.5	6.9	6.9	7.0	7.1	7.0	8.0	8.0	8.0	7.9	8.1
25	6.8	7.1	7.1	7.1	7.0	8.0	7.9	7.9	7.9	8.0
50	6.9	7.0	6.9	6.9	6.9	7.8	7.9	8.0	8.1	8.0
100	7.0	6.9	6.9	6.8	6.9	7.8	7.8	7.9	7.9	7.8

Table 2. Salinity and temperature in each concentration treatment in amphipod acute toxicity test.

Concentration treatment (%)		Sa	alinity (%	ю)			Tem	perature	(°C)		
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h	
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
6.5	30.0	30.0	30.0	30.0	30.0	0 22.0 22.0 22.0 22.0 22					
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	



Table 3. Ammonia-N, Total sulphide, Total suspended solids, Total residual chlorine concentration at the beginning and ending of the amphipod acute toxicity test.

Concentration treatment		onia-N L-1)		alphide L-1)	Total sus soli (mg	ds	Total re chlo (mg	
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.87	0.07	< 0.1	< 0.1	50	< 2	< 0.02	< 0.02
Salinity control	0.95	< 0.01	< 0.1	< 0.1	60	< 2	< 0.02	< 0.02
6.5	3.18	1.86	< 0.1	< 0.1	70	5	< 0.02	< 0.02
12.5	6.34	3.10	< 0.1	< 0.1	77	6	< 0.02	< 0.02
25	8.42	5.77	< 0.1	< 0.1	80	11	< 0.02	< 0.02
50	13.9	11.4	< 0.1	< 0.1	89	26	< 0.02	< 0.02
100	26.4	23.8	0.13	< 0.1	110	41	< 0.02	< 0.02



Appendix B

Monitoring Data for Fish Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in fish acute toxicity test.

Concentration treatment (%)		Dissolve	d oxygen	(mg L-1)				рН	_	
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	6.8	6.9	7.0	6.9	7.0	7.9	7.9	7.9	8.0	7.9
Salinity control	6.9	6.9	7.0	7.0	7.1	8.0	7.9	7.9	7.9	7.8
6.5	6.9	7.1	7.1	7.0	7.0	7.9	7.9	7.9	7.8	8.0
12.5	6.8	6.9	6.9	7.0	7.1	8.1	8.0	7.9	7.9	7.7
25	7.0	6.9	7.0	6.8	6.9	8.0	8.1	7.9	7.8	7.9
50	6.9	6.9	7.1	6.8	6.8	7.9	8.0	8.0	7.9	7.8
100	7.0	6.8	6.7	6.8	6.8	7.9	8.0	7.9	7.9	7.9

Table 2. Salinity and temperature in each concentration treatment in fish acute toxicity test.

Concentration treatment (%)		Sa	alinity (%	o)			Tem	perature	(°C)	
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, Total sulphide, Total suspended solids, Total residual chlorine concentration at the beginning and ending of the fish acute toxicity test.

Concentration treatment		onia-N ; L-1)	Total Sı (mg	ulphide (L-1)	Total sus soli (mg	ids	chlo	esidual vrine ; L-1)
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	< 0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
Salinity control	< 0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
6.5	1.95	1.45	< 0.1	< 0.1	3	2	< 0.02	< 0.02
12.5	3.09	3.06	< 0.1	< 0.1	6	< 2	< 0.02	< 0.02
25	6.07	5.42	< 0.1	< 0.1	10	3	< 0.02	< 0.02
50	12.9	10.3	< 0.1	< 0.1	16	6	< 0.02	< 0.02
100	23.6	23.0	0.19	< 0.1	41	8	< 0.02	< 0.02



Appendix C

Monitoring Data for Barnacle Larvae Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment (%)		Dissolve	d oxygen	(mg L-1)				рН		
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	6.8	6.9	6.9	6.9	6.8	7.9	8.0	8.0	7.9	7.8
Salinity control	6.9	6.9	7.0	7.0	6.8	7.9	8.0	8.0	7.8	7.8
6.5	6.9	7.0	6.8	6.9	6.8	8.0	7.9	7.9	7.9	8.0
12.5	6.9	6.9	6.9	6.8	6.8	7.9	7.9	7.9	7.9	8.0
25	6.9	6.9	6.8	6.9	7.0	7.9	7.8	7.8	7.8	7.8
50	6.8	6.9	6.9	6.9	7.1	7.9	7.9	7.9	7.8	7.7
100	6.8	6.8	6.8	6.9	6.8	7.8	7.8	7.8	7.8	7.7

Table 2. Salinity and temperature in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment (%)		Sa	alinity (%	o)			Tem	perature	(°C)	
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, Total sulphide, Total suspended solids, Total residual chlorine concentration at the beginning and ending of the barnacle larvae acute toxicity test.

01 1110 2 1111111		ore restrictly r	es 					_
Concentration	Ammo	onia-N	Total S	ulphide	Total susper	nded solids	Total residu	ual chlorine
treatment	(mg	; L-1)	(mg	(L-1)	(mg	L-1)	(mg	(L-1)
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
Salinity control	< 0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
6.5	1.23	0.97	< 0.1	< 0.1	3	< 2	< 0.02	< 0.02
12.5	2.53	2.00	< 0.1	< 0.1	4	< 2	< 0.02	< 0.02
25	5.83	4.44	< 0.1	< 0.1	11	< 2	< 0.02	< 0.02
50	10.5	8.76	< 0.1	< 0.1	17	< 2	< 0.02	< 0.02
100	22.9	17.4	0.24	< 0.1	41	3	< 0.02	< 0.02



Appendix D

Monitoring Data for Diatom Growth Inhibition Test (Chronic toxicity test)



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in diatom growth inhibition test.

Concentration			Disso	lved ox	ygen (n	ng L-1)]	рН			
treatment (%)	0 h	24 h	48 h	72 h	96 h	120h	144h	168h	0 h	24 h	48 h	72 h	96 h	120h	144h	168h
Negative control	6.8	6.9	7.1	7.2	7.5	7.7	8.0	8.2	8.0	7.9	8.0	8.0	7.9	8.1	8.1	8.1
Salinity control	6.9	6.8	7.0	7.2	7.4	7.8	7.9	8.2	7.9	7.9	8.1	8.0	8.0	8.1	8.0	8.1
2.5	6.9	6.9	7.2	7.2	7.6	7.8	8.1	8.2	8.0	7.9	7.9	8.0	8.1	8.1	8.1	8.0
5	6.8	6.8	7.1	7.1	7.6	7.9	8.0	8.2	8.0	8.0	8.1	8.0	7.9	7.9	8.0	8.1
10	6.8	6.9	7.1	7.3	7.4	7.6	7.9	8.3	7.9	7.9	8.0	8.1	8.0	8.0	8.0	8.1
25	6.9	6.8	7.0	7.1	7.2	7.6	7.8	8.2	7.9	8.0	8.0	8.0	8.1	8.0	7.9	8.0
50	6.8	6.8 6.9 7.0 7.2 7.4 7.4 7.6								7.9	7.9	8.0	8.0	8.0	8.1	8.0
100	6.9	6.8	6.9	6.9	6.9	7.1	7.2	7.3	7.9	7.8	7.9	7.9	7.9	7.9	8.0	8.0

Table 2. Salinity and temperature in each concentration treatment in diatom growth inhibition test.

Concentration treatment (%)				Salini	ty (‰)			J				Temper	rature (°	C)		
treatment (70)	0 h	24h	48h	72h	96h	120h	144h	168h	0h	24h	48h	72h	96h	120h	144h	168h
Negative control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
2.5	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
5	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
10	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, Total sulphide, Total suspended solids, Total residual chlorine concentration at the beginning and ending of the diatom growth inhibition toxicity test.

Concentration treatment		onia-N ; L-1)	Total Sı (mg	ılphide L-1)	Total sus soli (mg	ids	Total residu (mg	ual chlorine (L-1)
(%) -	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
Salinity control	< 0.01	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
2.5	2.49	< 0.01	0.12	< 0.1	3	2	< 0.02	< 0.02
5	2.33	< 0.01	0.16	< 0.1	6	4	< 0.02	< 0.02
10	5.40	< 0.01	0.18	< 0.1	9	5	< 0.02	< 0.02
25	7.72	< 0.01	0.22	< 0.1	15	10	< 0.02	< 0.02
50	11.2	5.09	0.23	< 0.1	19	14	< 0.02	< 0.02
100	22.4	19.5	0.26	0.24	47	30	< 0.02	< 0.02



Appendix E

Monitoring Data for Shrimp Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in shrimp acute toxicity test.

	0								J	
Concentration treatment (%)		Dissolve	d oxygen	(mg L-1)				рН		
treatment (70)	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	6.8	6.9	6.9	6.9	6.8	7.9	7.9	8.0	8.0	8.0
Salinity control	6.9	6.9	6.9	7.0	7.0	8.0	7.9	8.0	7.9	8.0
6.5	6.9	6.9	6.9	7.0	7.0	7.9	7.9	8.0	8.0	7.9
12.5	6.9	6.9	6.9	7.0	6.9	7.9	7.9	7.9	7.9	7.9
25	6.9	7.1	7.0	7.0	7.0	8.0	8.0	7.9	8.0	7.9
50	6.9	7.0	7.0	6.9	6.8	7.9	7.9	7.9	7.7	7.7
100	7.0	7.0	6.9	6.8	6.8	7.9	8.0	7.9	7.8	7.7

Table 2. Salinity and temperature in each concentration treatment in shrimp acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
Salinity control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
6.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
12.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
25	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
50	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
100	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, Total sulphide, Total suspended solids, Total residual chlorine concentration at the beginning and ending of the shrimp acute toxicity test.

	e originally the	tere territerly t	004.					
Concentration treatment (%)	Ammonia-N (mg L-1)		Total Sulphide (mg L-1)		Total suspended solids (mg L ⁻¹)		Total residual chlorine (mg L-1)	
	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	1.67	< 0.01	< 0.1	< 0.1	< 2	< 2	< 0.02	< 0.02
Salinity control	1.80	< 0.01	< 0.1	< 0.1	2	< 2	< 0.02	< 0.02
6.5	2.83	2.37	< 0.1	< 0.1	6	< 2	< 0.02	< 0.02
12.5	5.04	4.48	< 0.1	< 0.1	6	4	< 0.02	< 0.02
25	4.15	4.07	< 0.1	< 0.1	7	2	< 0.02	< 0.02
50	14.5	14.0	< 0.1	< 0.1	27	6	< 0.02	< 0.02
100	24.5	23.9	0.24	< 0.1	37	9	< 0.02	< 0.02