

Term Contract for Provision of Sampling and Analyzing of Wastewater and Sludge Samples for Various Sewage Treatment Facilities and Marine Water Samples in Urban Area, Lantau and Outlying Islands to the Drainage Services Department

Whole Effluent Toxicity Test (WETT) at SCISTW

Report for the Month of January 2021

Contract No. : DE/2020/02

Applicant : DRAINAGE SERVICES DEPT. - DIVISION 2

Address : STONECUTTERS ISLAND SEWAGE

TREATMENT WORKS, NGONG SHUEN CHAU,

KOWLOON, HONG KONG

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For and on behalf of

CMA Industrial Development Foundation Limited

Authorized Signature :

Lau Yan Kin Senior Manager Environmental Division



TESTING

Contents

1	Intr	oduction	4
	1.1	Background	4
	1.2	Testing laboratory and investigator	4
	1.3	Sample	4
	1.4	Test species	4
	1.5	Test protocols	5
2	Rep	ort on Amphipod Acute Toxicity Test	6
	2.1	Samples storage and pretreatment	7
	2.2	Test organism	7
	2.3	Summary of test conditions	7
	2.4	Test results	8-9
	2.5	Summary of water quality parameters monitoring during test	10
	2.6	LC ₅₀ for the amphipod Melita longidactyla and test acceptability	11
3	Rep	ort on Fish Acute Toxicity Test	12
	3.1	Samples storage and pretreatment	13
	3.2	Test organism	13
	3.3	Summary of test conditions	13
	3.4	Test results	14-16
	3.5	Summary of water quality parameters monitoring during test	16
	3.6	LC ₅₀ for the fish Lutjanus malabaricus and test acceptability	17
4	Rep	ort on Barnacle Larvae Acute Toxicity Test	18
	4.1	Samples storage and pretreatment	19
	4.2	Test organism	19
	4.3	Summary of test conditions	19
	4.4	Test results	20-21
	4.5	Summary of water quality parameters monitoring during test	22
	4.6	LC ₅₀ for the barnacle larvae <i>Balanus amphitrite</i> and test acceptability	23
5	Rep	ort on Diatom Growth Inhibition Test (Chronic toxicity test)	24
	5.1	Samples storage and pretreatment	25
	5.2	Test organism	25
	5.3	Summary of test conditions	25
	5.4	Test results	26-27
	5.5	Summary of water quality parameters monitoring during test	28
	5.6	IC50 for the diatom Skeletonema costatum and test acceptability	29
6	Rep	ort on Shrimp Acute Toxicity Test	30
	6.1	Samples storage and pretreatment	31
	6.2	Test organism	31
	6.3	Summary of test conditions	31
	6.4	Test results	32-33
	6.5	Summary of water quality parameters monitoring during test	34
	6.6	LC ₅₀ for the shrimp <i>Metapenaeus ensis</i> and test acceptability	35



7 Conclusion	36-37
Appendix A Monitoring Data for Amphipod Acute Toxicity Test	38-41
Appendix B Monitoring Data for Fish Acute Toxicity Test	42-45
Appendix C Monitoring Data for Barnacle Larvae Acute Toxicity Test	46-49
Appendix D Monitoring Data for Diatom Growth Inhibition Test	50-53
Appendix E Monitoring Data for Shrimp Acute Toxicity Test	54-57



1. Introduction

1.1. Background

The whole effluent toxicity tests (WETT) were carried out under the requirements of Drainage Service Department (DSD).

1.2. <u>Testing laboratory and investigator</u>

Principle investigator: Prof. Wen-Xiong WANG

Phone number: (852) 3442-4693

Address: School of Energy and Environment, City Univ. Hong Kong, Hong Kong

1.3. Sample

A 24-hour flow-weighted composite effluent sample was collected from Stonecutters Island Sewage Treatment Works (SCISTW) on January 15, 2021. Effluent sample was shipped immediately to the testing laboratory on the same day of collection and stored at 4 °C until use. Toxicity testings started on Day 6 after the sample collection.

Toxicity testings started on Day 6 after sample collection was due to the delayed shipment of tested fish to the testing laboratory caused by COVID-19 situation. The storage of effluents for a few days should not affect the overall testing results for the following reasons:

- 1) The samples were stored at low temperature (4 °C) and under dark conditions; thus there should be minimal biological activity (if any) at such low temperature;
- 2) The effluents were also stored under sealed conditions; thus there should be minimal loss of chemicals (or contaminants) from the water to the air, i.e., evaporation loss;
- 3) All the stored containers were in polycarbonate quality, and there should be minimal absorption of chemicals to the container walls during the storage periods.

Thus, with minimal biological activity and chemical loss (either evaporation or adsorption), these effluents should keep stable within the few days of storage. The toxicity testing results should reflect the conditions when the effluents were originally collected.

1.4. <u>Test species</u>

The following test species were included in the WETT:

- Amphipod (Melita longidactyla)
- Fish (*Lutjanus malabaricus*)
- Barnacle larvae (*Balanus amphitrite*)
- Diatom (*Skeletonema costatum*)
- Shrimp (*Metapenaeus ensis*)



1.5. Test protocols

The WETT testing methods and procedures follow those documented in "Consultancy Study on Fisheries and Marine Ecological Criteria for Impact Assessment-Final Report" commissioned by Agriculture, Fisheries and Conservation Department (AFCD), as indicated in tender addendum No. 1 by Drainage Services Department (DSD).



2. Report on Amphipod Acute Toxicity Test



Test report

2.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

2.2. <u>Test organism</u>

Species Amphipod (Melita longidactyla).

Source: Collected from local coastal waters from Sai Kung

Size/age: 0.5-0.7 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature: 22±1°C,

salinity: 30‰) at least 48 hours in the laboratory prior to test.

Fed with green algae Ulva lactuca.

2.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 21/01/2021-23/01/2021

Dilution seawater source: Seawater collected from a pristine site in Clear Water Bay, Sai

Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

Testing temperature: 22±1 °C
Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 150 mL glass flask

Feeding: None
Number of organisms per replicate: 10
Replicate number: 4

Volume of test medium: 100 mL

Aeration: Moderate, around 100 bubbles/min

Reference toxicant: CdCl₂

Positive control: 48-h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



2.4. Test results

Table 1. Survival of amphipods after 48 hours.

Treatment	Effluent	Number of living amphipods after 48 hour (individuals)					
rreatment	concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	9	10	8	10	9.25	0.96
Salinity control	0	8	9	8	10	8.75	0.96
Concentration 1	6.5	10	9	10	9	9.50	0.58
Concentration 2	12.5	6	8	7	6	6.75	0.96
Concentration 3	25	6	6	4	5	5.25	0.96
Concentration 4	50	1	2	1	0	1.00	0.82
Concentration 5	100	0	0	0	0	0.00	0.00



Table 2. Survival percentage of amphipods after 48 hours.

Treatment	Effluent concentration	Percentage of living amphipods after 48 hour (%)						
Treatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	90	100	80	100	92.50	9.57	
Salinity control	0	80	90	80	100	87.50	9.57	
Concentration 1	6.5	100	90	100	90	95.00	5.77	
Concentration 2	12.5	60	80	70	60	67.50	9.57	
Concentration 3	25	60	60	40	50	52.50	9.57	
Concentration 4	50	10	20	10	0	10.00	8.16	
Concentration 5	100	0	0	0	0	0.00	0.00	



2.5 Summary of water quality parameters monitoring during test.

Table 3. Summary of water quality parameters during amphipod acute toxicity test.

		Effluent concentration (%)							
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100		
Salinity (%)	30.0	30.0	30.0	30.0	30.0	30.0	30.0		
Dissolved oxygen (mg L-1)	6.8-7.7	6.8-7.1	6.9-7.1	6.9-7.1	6.8-7.1	6.9-7.0	6.8-6.9		
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0		
рН	7.9-8.1	7.9-8.1	7.9-8.1	8.0-8.1	7.9-8.0	7.8-8.0	7.8-7.9		
Total ammonia (start/end, mg L-1)	0.03/1.36	0.04/2.20	1.17/2.12	2.78/4.61	6.57/11.3	13.2/14.7	26.4/20.9		
Total sulfide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	<0.1		
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02		
Total suspended solid (start/end, mg L-1)	<2	<2	<2	<2	<2	<2	<2		



2.5. LC₅₀ for the amphipod *Melita longidactyla* and test acceptability

Table 4. LC₅₀ for the amphipods and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	23.07 %	NA
Negative control survival	92.50%	>90%
Reference toxicant 48-h acute test	1.34 mg L ⁻¹	1.25±0.15 mg L ⁻¹
95% of confidence range of reference toxicant test	1.09-1.56 mg L ⁻¹	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: $\pm 1~^{\circ}\text{C}$
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L ⁻¹

NA: Not applicable



3. Report on Fish Acute Toxicity Test



Test report

3.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

3.2. <u>Test organism</u>

Species Fish (Lutjanus malabaricus)

Source: Purchased from local contracted fish farm

Size/age: 2-3 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature: 22±1°C,

salinity: 30%) at least 48 hours in laboratory prior to test. Fed

with fresh shrimp purchased from local market.

3.3. <u>Summary of test conditions</u>

Type of test: Static

Duration: 48 h, 21/01/2021-23/01/2021

Dilution seawater source: Seawater collected from a pristine site in Clear Water Bay, Sai

Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 5 μm filtration bag

Testing temperature: 22±1 °C Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 20 L tank

Feeding: None
Number of organisms per replicate: 20
Replicate number: 4
Volume of test medium: 20 L

Aeration: Moderate, with air stone

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



3.4. Test results

T	F(G		Numbe	er of living fish af	ter 48 hour (indivi	duals)	
Treatment	Effluent concentration (%) -	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	20	19	19	20	19.50	0.58
Salinity control	0	18	20	20	19	19.25	0.96
Concentration 1	6.5	17	19	18	19	18.25	0.96
Concentration 2	12.5	19	17	18	18	18.00	0.82
Concentration 3	25	17	16	15	14	15.50	1.29
Concentration 4	50	8	7	8	9	8.00	0.82
Concentration 5	100	0	1	1	0	0.50	0.58

Table 1. Survival of fish after 48 hours.



Table 2. Survival percentage of fish after 48 hours.

Treatment	Effluent concentration (%)		Per	centage of living	fish after 48 hour ('	%)						
rreatment	Enfuent Concentration (%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD					
Negative control	0	100	95	95	100	97.50	2.89					
Salinity control	0	90	100	100	95	96.25	4.79					
Concentration 1	6.5	85	95	90	95	91.25	4.79					
Concentration 2	12.5	95	85	90	90	90.00	4.08					
Concentration 3	25	85	80	75	70	77.50	6.45					
Concentration 4	50	40	35	40	45	40.00	4.08					
Concentration 5	100	0	5	5	0	2.50	2.89					



3.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during fish acute toxicity test.

	Effluent concentration (%)								
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100		
Salinity (‰)	30	30	30	30	30	30	30		
Dissolved oxygen (mg L-1)	6.8-7.0	6.9-7.7	6.9-7.1	6.8-7.0	6.8-7.0	6.8-7.1	6.7-7.0		
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0		
pH	7.9-8.0	7.9-8.0	7.9-8.0	7.7-8.0	7.8-8.1	7.8-8.0	7.9-8.0		
Total ammonia (start/end, mg L-1)	3.11/2.82	2.44/3.01	4.53/6.03	7.62/6.85	7.86/7.60	16.4/16.7	26.0/25.4		
Total sulfide (start/end, mg L-1)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1		
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02		
Total suspended solid (mg L-1)	4/3	3/3	5/4	5/7	13/13	26/25	47/37		



3.6. LC_{50} for the fish *Lutjanus malabaricus* and test acceptability

Table 4. LC₅₀ for the fish and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	42.70 %	NA
Negative survival	97.50 %	>90%
Reference toxicant 48-h acute test	13.05 mg L ⁻¹	14.6±1.78 mg L ⁻¹
95% of confidence range of reference toxicant test	$11.54-15.19~{ m mg}~{ m L}^{-1}$	NA
Daily temperature variation	<0.5 °C	Average daily temperature variation: $\pm1^{\circ}\text{C}$
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L ⁻¹

a: The mortalities in all concentration groups were less than 50% of that in control group and thus LC₅₀ cannot be calculated.

calculated.

NA: Not applicable



4. Report on Barnacle Larvae Acute Toxicity Test



Test report

4.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 μ m membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

4.2. <u>Test organism</u>

Species Barnacle larvae (Balanus amphitrite).

Source: Introduced from adult barnacles collected from Sai Kung

Size/age: Stage II

Acclimatization: Acclimatized in fully aerated seawater held in 500 mL glass

beaker (temperature: 22±1°C, salinity: 30‰) for at least 24 hours in laboratory prior to test. Fed with diatom *Chaetoceros*

gracilis.

4.3. Summary of test conditions

Type of test: Static

Duration: 48 h, 21/01/2021-23/01/2021

Dilution seawater source: Seawater collected from a pristine site in Clear Water Bay, Sai

Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 µm membrane

Testing temperature: $22\pm1\,^{\circ}\text{C}$ Lighting: Continuous

Salinity: 30%

Testing chamber: Pre-cleaned 50 mL glass beaker

Feeding: None
Number of organisms per replicate: 20
Replicate number: 4
Volume of test medium: 20 mI

Aeration: Moderate, around 100 bubbles/min

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 30‰



4.4. <u>Test results</u>

Table 1. Survival of barnacle larvae after 48 hours

Treatment	Effluent concentration	Number of living barnacle larvae after 48 hour (individuals)					
Treatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	20	20	20	20	20.00	0.00
Salinity control	0	19	18	17	18	18.00	0.82
Concentration 1	6.5	20	16	17	16	17.25	1.89
Concentration 2	12.5	16	16	17	16	16.25	0.50
Concentration 3	25	13	14	13	9	12.25	2.22
Concentration 4	50	15	9	11	14	12.25	2.75
Concentration 5	100	0	0	0	0	0.00	0.00



Table 2. Survival percentage of barnacle larvae after 48 hours.

T	Effluent concentration		Percentag	ge of living barna	cle larvae after 48	hour (%)	
Treatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	100	100	100	100	100.00	0.00
Salinity control	0	95	90	85	90	90.00	4.08
Concentration 1	6.5	100	80	85	80	86.25	9.46
Concentration 2	12.5	80	80	85	80	81.25	2.50
Concentration 3	25	65	70	65	45	61.25	11.09
Concentration 4	50	75	45	55	70	61.25	13.77
Concentration 5	100	0	0	0	0	0.00	0.00



4.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during barnacle larvae acute toxicity test

		0						
	Effluent concentration (%)							
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100	
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0	
Dissolved oxygen (mg L-1)	6.8-6.9	6.8-7.0	6.8-7.0	6.8-6.9	6.8-7.0	6.8-7.1	6.8-6.9	
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0	
pH	7.8-8.0	7.8-8.0	7.9-8.0	7.9-8.0	7.8-7.9	7.7-7.9	7.7-7.8	
Total ammonia (start/end, mg L-1)	0.04/0.87	0.03/0.92	0.04/1.43	0.05/1.57	0.07/1.80	0.08/2.63	0.20/6.58	
Total sulfide (start/end, mg L-1)	<0.1	< 0.1	<0.1	<0.1	< 0.1	< 0.1	< 0.1	
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	
Total suspended solid (mg L-1)	<2	<2	<2	<2	<2	<2	<2	



4.6. LC₅₀ for the barnacle larvae Balanus amphitrite and test acceptability

Table 4. LC_{50} for the barnacle larvae and test acceptability

Parameter	Value	Control limit
Calculated LC ₅₀	33.67 %	NA
Negative survival	100.00 %	>90%
Reference toxicant 48-h acute test	0.99 mg L^{-1}	1.04±0.11 mg L ⁻¹
95% of confidence range of reference toxicant test:	$0.87 \text{-} 1.23 \text{ mg L}^{-1}$	NA
Daily temperature variation	<0.5℃	Average daily temperature variation: $\pm1^{\rm o}{\rm C}$
Dissolved oxygen concentration	>6.7 mg L ⁻¹	>4 mg L ⁻¹

NA: Not applicable



5. Report on Diatom Growth Inhibition Test (Chronic toxicity test)



Test report

5.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 5 μ m membrane filter to remove large debris. Effluent was added with ocean salt in order to raise the salinity to the required level (30‰) and then aerated moderately to dissolved oxygen (DO) saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

5.2. <u>Test organism</u>

Species Diatom (Skeletonema costatum)

Source: Grown from laboratory culture obtained from Coastal Marine

Lab, Hong Kong University of Science and Technology

Size/age: Log growth phase

Acclimatization: Grown in 250 mL glass flask (temperature: 22±1°C, salinity:

30‰, 3000 lux) for at least two weeks prior to test.

5.3. Summary of test conditions

Type of test: Static

Duration: 7 days, 21/01/2021-28/01/2021

Dilution seawater source: Seawater collected from a pristine site in Clear Water Bay, Sai

Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

Testing temperature: 22±1 °C

Lighting: 12 h light/12 h dark cycle, 3000±500 lux

Salinity: 30%

Testing chamber: Pre-cleaned 100 mL glass beaker

Initial cell density: $(5.0\pm0.4)\times10^4$ cell mL⁻¹

Replicate number: 4

Volume of test medium: 25 mL

Aeration: None

Reference toxicant: CdCl2

Positive control: 7-day IC₅₀ toxicity test

Salinity control:

Prepared with ocean salt adding into de-ionized water,

salinity: 30%



5.4. <u>Test results</u>

Table 1. Cell density of diatom *Skeletonema costatum* at the beginning and end of growth inhibition test. Initial cell density: (5.0±0.4)×10⁴ cell mL⁻¹.

Treatment	Effluent concentration	Cell density after 7-day growth (×106 cell mL-1)						
rreatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	1.12	1.10	1.13	1.12	1.12	0.01	
Salinity control	0	1.13	1.12	1.13	1.15	1.13	0.01	
Concentration 1	2.5	1.21	1.18	1.20	1.24	1.21	0.03	
Concentration 2	5	1.88	1.94	1.97	1.85	1.91	0.05	
Concentration 3	10	1.53	1.44	1.53	1.52	1.51	0.04	
Concentration 4	25	0.9	1.01	1.07	0.96	0.99	0.07	
Concentration 5	50	0.07	0.08	0.04	0.05	0.06	0.02	
Concentration 6	100	0	0	0	0	0.00	0.00	



Table 2. Growth rate of *Skeletonema costatum* within 7 days.

Treatment	Effluent concentration	Fluent concentration 7-day average growth rate (d-1)					
Treatment	eatment (%)		Replicate 2	Replicate 3	Replicate 4	Mean	SD
Negative control	0	0.44	0.44	0.45	0.44	0.44	0.00
Salinity control	0	0.45	0.44	0.45	0.45	0.45	0.00
Concentration 1	2.5	0.46	0.45	0.45	0.46	0.45	0.00
Concentration 2	5	0.52	0.52	0.52	0.52	0.52	0.00
Concentration 3	10	0.49	0.48	0.49	0.49	0.49	0.00
Concentration 4	25	0.41	0.43	0.44	0.42	0.43	0.01
Concentration 5	50	0.05	0.07	-0.03	0.00	0.02	0.05
Concentration 6	100	0.00	0.00	0.00	0.00	0.00	0.00



5.5. Summary of water quality parameters monitoring during test

Table 3. Summary of water quality parameters during diatom growth inhibition test

	Effluent concentration (%)							
Water quality parameters	Negative control	Salinity control	2.5	5.0	10	25	50	100
Salinity (‰)	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0
Dissolved oxygen (mg L-1)	6.8-8.2	6.8-8.3	6.8-8.2	6.8-8.2	6.8-8.3	6.8-8.2	6.8-7.9	6.8-7.3
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
pН	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.1	7.9-8.0
Total ammonia (start/end, mg L-1)	<0.01/<0.01	<0.01/<0.01	1.27/1.31	2.70/2.86	4.12/4.37	6.22/6.43	11.2/9.86	25.3/24.5
Total sulfide (start/end, mg L-1)	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1	< 0.1
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02
Total suspended solid (mg L-1)	<2	<2	<2	<2	<2	<2	<2	<2



5.6. IC₅₀ for the diatom Skeletonema costatum and test acceptability

Table 4. IC₅₀, none observed effect concentration (NOEC) for the diatom and test acceptability

Parameter	Value	Control limit
Calculated IC ₅₀	32.23 %	NA
None observed effect concentration (NOEC)	10 %	-
Reference toxicant 7-day test:	$0.11~{ m mg~L^{-1}}$	$0.13\pm0.02~{ m mg}~{ m L}^{-1}$
95% of confidence range of reference toxicant test	0.09-0.15 mg L-1	NA
Temperature variation	<0.5 °C	Average daily temperature variation: $\pm1^{\circ}\text{C}$

NA: Not applicable



6. Report on Shrimp Acute Toxicity Test



Test report

6.1. Samples storage and pretreatment

Effluent sample was thoroughly mixed and passed through 2-mm mesh to remove the large debris. Effluent was added with ocean salt in order to raise the salinity to 25% and then aerated moderately such that the dissolved oxygen (DO) reached saturation prior to use. Salinity control was set up to monitor if there was adverse effect on the test organisms.

6.2. <u>Test organism</u>

Species Shrimp (Metapenaeus ensis).

Source: Purchased from contracted fish dealer

Size/age: 5-7 cm

Acclimatization: Acclimatized in fully aerated seawater (temperature: 22±1°C,

salinity: 25%) at least 48 hours in the laboratory prior to test.

Fed with commercial shrimp feeds.

6.3. Summary of test conditions

Type of test: Static

Duration:

48 h, 21/01/2021-23/01/2021

Dilution seawater source: Seawater collected from a pristine site in Clear Water Bay, Sai

Kung, Hong Kong

Dilution seawater pretreatment: Filtered through 0.22 μm membrane

Testing temperature: $22\pm1\,^{\circ}\text{C}$ Lighting: Continuous

Salinity: 25‰

Testing chamber: Pre-cleaned 20 L tank

Feeding: None
Number of organisms per replicate: 10
Replicate number: 4
Volume of test medium: 10 L

Aeration: Moderate, with air stone

Reference toxicant: CdCl₂

Positive control: 48 h acute toxicity test

Salinity control: Prepared with ocean salt adding into de-ionized water,

salinity: 25‰



6.4. <u>Test results</u>

Table 1. Survival of shrimps after 48 hours.

T	Effluent concentration	Number of living shrimps after 48 hour (individuals)						
Treatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	10	9	9	10	9.50	0.58	
Salinity control	0	9	8	10	10	9.25	0.96	
Concentration 1	6.5	8	7	8	7	7.50	0.58	
Concentration 2	12.5	7	7	6	6	6.50	0.58	
Concentration 3	25	7	7	5	6	6.25	0.96	
Concentration 4	50	4	5	4	4	4.25	0.50	
Concentration 5	100	1	0	0	0	0.25	0.50	



Table 2. Survival percentage of shrimps after 48 hours.

Torontoront	Effluent concentration	Percentage of living shrimps after 48 hour (%)						
Treatment	(%)	Replicate 1	Replicate 2	Replicate 3	Replicate 4	Mean	SD	
Negative control	0	100	90	90	100	95.00	5.77	
Salinity control	0	90	80	100	100	92.50	9.57	
Concentration 1	6.5	80	70	80	70	75.00	5.77	
Concentration 2	12.5	70	70	60	60	65.00	5.77	
Concentration 3	25	70	70	50	60	62.50	9.57	
Concentration 4	50	40	50	40	40	42.50	5.00	
Concentration 5	100	10	0	0	0	2.50	5.00	



6.5. Summary of water quality parameters monitoring during test.

Table 3. Summary of water quality parameters during shrimp acute toxicity test.

	Effluent concentration (%)							
Water quality parameters	Negative control	Salinity control	6.5	12.5	25	50	100	
Salinity (‰)	25.0	25.0	25.0	25.0	25.0	25.0	25.0	
Dissolved oxygen (mg L-1)	6.8-6.9	6.9-7.0	6.9-7.0	6.9-7.0	6.9-7.1	6.8-7.0	6.8-7.0	
Temperature (°C)	22.0	22.0	22.0	22.0	22.0	22.0	22.0	
рН	7.9-8.0	7.9-8.0	7.9-8.0	7.9-7.9	7.9-8.0	7.7-7.9	7.7-8.0	
Total ammonia (start/end, mg L-1)	0.05/0.05	1.26/1.56	1.43/1.84	3.13/3.42	6.03/7.25	15.4/14.4	28.2/27.0	
Total sulfide (start/end, mg L-1)	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
Total residual chlorine (start/end, mg L-1)	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	< 0.02	
Total suspended solid (start/end, mg L-1)	4/5	4/6	5/6	6/9	24/22	45/32	50/62	



6.6 LC₅₀ for the shrimp Metapenaeus ensis and test acceptability

Table 4. LC₅₀ for the *Metapenaeus ensis* and test acceptability.

Parameter	Value	Control limit
Calculated LC ₅₀	28.57 %	NA
Negative control survival	95.00 %	>90%
Reference toxicant 48-h acute test	5.62 mg L ⁻¹	5.19±0.51 mg L ⁻¹
95% of confidence range of reference toxicant test	4.83-7.02 mg L ⁻¹	NA
Daily temperature variation	<0.5°C	Average daily temperature variation: $\pm 1~^{\circ}\text{C}$
Dissolved oxygen concentration	>6.7 mg L-1	>4 mg L ⁻¹

NA: Not applicable



7. Conclusion



Table 1. Comparison of measured toxicity values with the target toxicity levels.

Test species	Measured LC ₅₀ /IC ₅₀ /NOEC	Target toxicity level
Amphipod Melita longidactyla	23.07 %	≥7.1%
Fish Lutjanus malabaricus	42.70 %	≥7.1%
Barnacle larvae Balanus amphitrite	33.67 %	≥7.1%
Diatom Skeletonema costatum	32.23 %	-
Diatom Skeletonema costatum	10 % (NOEC)	≥0.51%
Shrimp Metapenaeus ensis	28.57 %	≥7.1%

Conclusion: all the measured values met the target toxicity levels as indicated in the EM&A Manual.



Appendix A

Monitoring Data for Amphipod Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in amphipod acute toxicity test.

Concentration treatment (%)		Dissol	ved oxygen (:	mg L ⁻¹)				рН				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h		
Negative control	6.8	6.9	7.0	7.0	7.1	8.0	7.9	8.1	8.1	8.0		
Salinity control	6.8	6.9	7.1	7.1	7.0	7.9	8.0	7.9	8.0	8.1		
6.5	6.9	7.0	7.1	7.0	7.1	8.0 7.9 8.0 8.1						
12.5	6.9	6.9	7.1	7.0	7.0	8.0	8.0	8.0	8.0	8.1		
25	6.8	7.0	7.1	7.1	7.0	8.0	8.0	8.0	7.9	8.0		
50	6.9	7.0	6.9	6.9	7.0	7.8	7.9	8.0	8.0	8.0		
100	6.9	6.9	6.9	6.8	6.9	7.8	7.8	7.9	7.8	7.8		



Table 2. Salinity and temperature in each concentration treatment in amphipod acute toxicity test.

Concentration treatment (%)		9	Salinity (‰)	,		Ten	nperature	(°C)	
	0 h	12 h	24 h	36 h	0 h	12 h	24 h	36 h	48 h	
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0



Table 3. <u>Ammonia-N</u>, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the amphipod acute toxicity test.

Concentration treatment	Ammonia (mg L-1)		_	hide L-1)	Total suspe (mg	nded solids L-1)	Total residual chlorine (mg L-1)		
(%)	Initial	End	Initial	End	Initial	End	Initial	End	
Negative control	0.03	1.36	<0.1	<0.1	<2	<2	<0.02	<0.02	
Salinity control	0.04	2.20	<0.1	<0.1	<2	<2	< 0.02	< 0.02	
6.5	1.17	2.12	<0.1	<0.1	<2	<2	< 0.02	< 0.02	
12.5	2.78	4.61	<0.1	<0.1	<2	<2	< 0.02	< 0.02	
25	6.57	11.3	<0.1	<0.1	<2	<2	< 0.02	< 0.02	
50	1.32	14.7	<0.1	<0.1	<2	<2	< 0.02	< 0.02	
100	26.4	20.9	<0.1	<0.1	<2	<2	< 0.02	< 0.02	



Appendix B

Monitoring Data for Fish Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in fish acute toxicity test.

Table 1. Bissorved oxygen		centration and print each concentration freatment in fibrial acute toxicity test.									
Concentration treatment (%)		Dissolv	ed oxygen ((mg L-1)			рН				
	0 h	12 h	24 h	36 h	0 h	12 h	24 h	36 h	48 h		
Negative control	6.8	6.8	6.9	6.9	7.0	7.9	7.9	7.9	8.0	7.9	
Salinity control	6.9	6.9	6.9	7.0	7.1	8.0	7.9	7.9	7.9	7.9	
6.5	6.9	7.0	7.1	6.9	7.0	7.9	7.9	7.9	7.8	8.0	
12.5	6.8	6.9	6.9	6.9	7.0	7.9	8.0	7.9	7.9	7.7	
25	7.0	6.9	7.0	6.8	6.9	8.0	8.1	7.9	7.8	7.9	
50	6.9	6.9	7.1	6.8	6.8	7.9	8.0	8.0	8.0	7.8	
100	7.0	6.7	6.8	6.8	6.8	7.9	8.0	7.9	7.9	7.9	



Table 2. Salinity and temperature in each concentration treatment in fish acute toxicity test.

Concentration treatment (%)			Salinity (‰)			Temperature (°C)					
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h	
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	



Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the fish acute toxicity test.

Concentration treatment	Ammo (mg		Sulp (mg		Total suspe (mg		Total residual chlorine (mg L ⁻¹)		
(%)	Initial	End	Initial	End	Initial	End	Initial	End	
Negative control	3.11	2.82	<0.1	<0.1	4	3	<0.02	<0.02	
Salinity control	2.44	3.01	<0.1	<0.1	3	3	< 0.02	< 0.02	
6.5	4.53	6.03	<0.1	<0.1	5	4	< 0.02	< 0.02	
12.5	7.62	6.85	<0.1	<0.1	5	7	< 0.02	< 0.02	
25	7.86	7.60	<0.1	<0.1	13	13	< 0.02	< 0.02	
50	16.4	16.7	<0.1	<0.1	26	25	< 0.02	< 0.02	
100	26.0	25.4	<0.1	<0.1	47	37	< 0.02	< 0.02	



Appendix C

Monitoring Data for Barnacle Larvae Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment (%)		Dissolv	ved oxygen (mg L ⁻¹)			рН			
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	6.8	6.8	6.9	6.9	6.8	7.9	7.9	8.0	7.9	7.8
Salinity control	6.9	6.8	6.9	7.0	6.8	7.9	7.9	8.0	7.8	7.8
6.5	6.9	7.0	6.8	6.8	6.9	8.0	8.0	7.9	7.9	8.0
12.5	6.9	6.8	6.9	6.9	6.8	7.9	7.9	7.9	7.9	8.0
25	6.9	6.9	6.8	6.9	7.0	7.9	7.8	7.8	7.8	7.8
50	6.9	6.8	6.9	6.9	7.1	7.9	7.8	7.9	7.8	7.7
100	6.8	6.8	6.8	6.9	6.8	7.8	7.8	7.8	7.8	7.7



Table 2. Salinity and temperature in each concentration treatment in barnacle larvae acute toxicity test.

Concentration treatment			Salinity (‰))			Ter	nperature	(°C)	
(%)	0 h	12 h	24 h	36 h	0 h	12 h	24 h	36 h	48 h	
Negative control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
6.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
12.5	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the barnacle larvae acute toxicity test.

Concentration treatment		onia-N (L-1)	Sulp (mg		Total suspe (mg			ıal chlorine L ⁻¹)
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.04	0.87	<0.1	<0.1	<2	<2	<0.02	<0.02
Salinity control	0.03	0.92	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
6.5	0.04	1.43	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
12.5	0.05	1.57	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
25	0.07	1.80	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
50	0.08	2.63	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
100	0.20	6.58	<0.1	< 0.1	<2	<2	< 0.02	< 0.02



Appendix D

Monitoring Data for Diatom Growth Inhibition Test (Chronic toxicity test)



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in diatom growth inhibition test.

Concentration			Disa	solved o	oxygen (mg L-1)							рН			
treatment (%)	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h	0 h	24 h	48 h	72 h	96 h	120 h	144 h	168 h
Negative control	6.8	6.9	7.1	7.2	7.5	7.7	8.0	8.2	8.0	7.9	8.0	8.0	7.9	8.1	8.1	8.1
Salinity control	6.9	6.8	7.0	7.2	7.4	7.8	7.9	8.3	7.9	7.9	8.1	8.0	8.0	8.1	8.0	8.1
2.5	6.9	6.9	7.2	7.2	7.6	7.8	8.1	8.2	8.0	7.9	7.9	8.0	8.1	8.1	8.1	8.0
5	6.8	6.8	7.1	7.1	7.6	7.9	8.0	8.2	8.0	8.0	8.1	8.0	7.9	7.9	8.0	8.1
10	6.8	6.9	7.1	7.3	7.4	7.6	7.9	8.3	7.9	7.9	8.0	8.1	8.0	8.0	8.0	8.1
25	6.9	6.8	7.0	7.1	7.2	7.6	7.8	8.2	7.9	8.0	8.0	8.0	8.1	8.0	7.9	8.0
50	6.8	6.9	7.0	7.2	7.4	7.4	7.6	7.8	7.9	7.9	7.9	8.0	8.0	8.0	8.1	8.0
100	6.9	6.8	6.9	6.9	6.9	7.1	7.2	7.3	7.9	7.8	7.9	7.9	7.9	7.9	8.0	8.0



Table 2. Salinity and temperature in each concentration treatment in diatom growth inhibition test.

Concentration				Salini	ty (‰)							Temper	rature (ºº	C)		
treatment (%)	0h	24h	48h	72h	96h	120h	144h	168h	0h	24h	48h	72h	96h	120h	144h	168h
Negative control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
Salinity control	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
2.5	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
5	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
10	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
25	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
50	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0
100	30.0	30.0	30.0	30.0	30.0	30.0	30.0	30.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the diatom growth inhibition toxicity test.

Concentration treatment	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspended solids (mg L-1)		Total residual chlorine (mg L ⁻¹)	
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	<0.01	<0.01	<0.1	<0.1	<2	<2	<0.02	<0.02
Salinity control	< 0.01	< 0.01	< 0.1	< 0.1	<2	<2	< 0.02	< 0.02
2.5	1.27	1.31	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
5	2.70	2.86	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
10	4.12	4.37	<0.1	< 0.1	<2	<2	< 0.02	< 0.02
25	6.22	6.43	< 0.1	< 0.1	<2	<2	< 0.02	< 0.02
50	11.2	9.86	< 0.1	< 0.1	<2	<2	< 0.02	< 0.02
100	25.3	24.5	< 0.1	< 0.1	<2	<2	< 0.02	< 0.02



Appendix E

Monitoring Data for Shrimp Acute Toxicity Test



Table 1. Dissolved oxygen concentration and pH in each concentration treatment in shrimp acute toxicity test.

Concentration treatment (%)		Dissolved oxygen (mg L-1)					рН					
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h		
Negative control	6.8	6.9	6.9	6.9	6.8	7.9	7.9	8.0	8.0	8.0		
Salinity control	6.9	6.9	6.9	6.9	7.0	8.0	7.9	8.0	7.9	8.0		
6.5	6.9	6.9	6.9	7.0	7.0	7.9	7.9	8.0	7.9	7.9		
12.5	6.9	6.9	6.9	7.0	6.9	7.9	7.9	7.9	7.9	7.9		
25	6.9	7.1	7.0	7.0	7.0	8.0	8.0	7.9	8.0	7.9		
50	6.9	7.0	7.0	6.9	6.8	7.9	7.9	7.9	7.7	7.7		
100	7.0	7.0	6.9	6.8	6.8	7.9	8.0	7.9	7.8	7.7		



Table 2. Salinity and temperature in each concentration treatment in shrimp acute toxicity test.

Concentration treatment (%)	Salinity (‰)					Temperature (°C)				
	0 h	12 h	24 h	36 h	48 h	0 h	12 h	24 h	36 h	48 h
Negative control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
Salinity control	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
6.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
12.5	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
25	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
50	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0
100	25.0	25.0	25.0	25.0	25.0	22.0	22.0	22.0	22.0	22.0



Table 3. Ammonia-N, sulphide, total suspended solids, total residual chlorine concentration at the beginning and ending of the shrimp acute toxicity test.

Concentration treatment	Ammonia-N (mg L ⁻¹)		Sulphide (mg L ⁻¹)		Total suspe (mg		Total residual chlorine (mg L-1)	
(%)	Initial	End	Initial	End	Initial	End	Initial	End
Negative control	0.05	0.05	<0.1	<0.1	4	5	<0.02	<0.02
Salinity control	1.26	1.56	<0.1	<0.1	4	6	< 0.02	< 0.02
6.5	1.43	1.84	<0.1	<0.1	5	6	< 0.02	< 0.02
12.5	3.13	3.42	<0.1	<0.1	6	9	< 0.02	< 0.02
25	6.03	7.25	<0.1	<0.1	24	22	< 0.02	< 0.02
50	15.4	14.4	<0.1	<0.1	45	32	< 0.02	< 0.02
100	28.2	27.0	<0.1	<0.1	50	62	< 0.02	< 0.02